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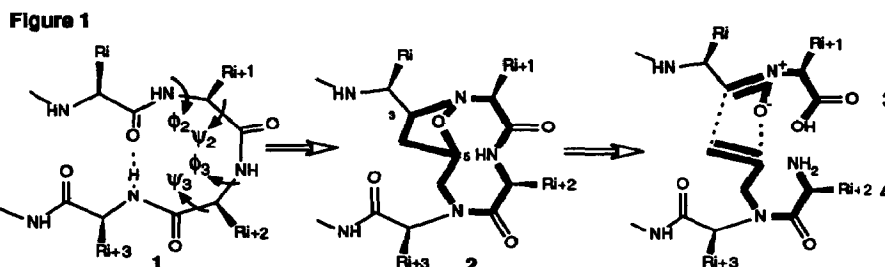
PEPTIDE BACKBONE-TO-BACKBONE CYCLISATION AS AN AVENUE TO
 β -TURN MIMICS

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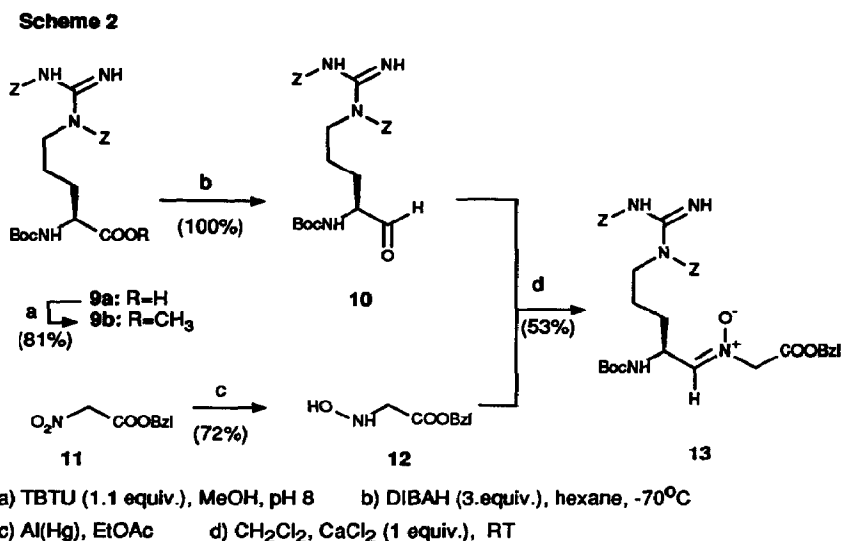
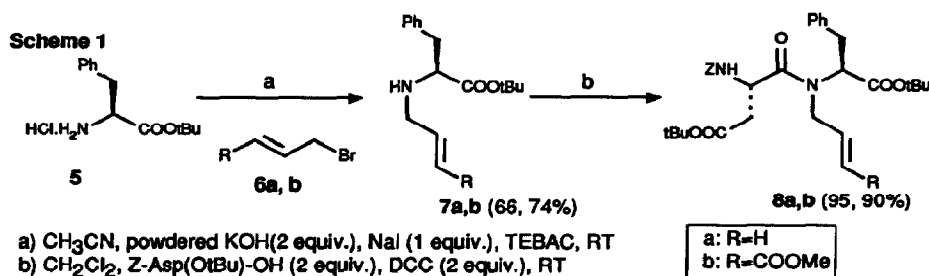
Abstract: 1,3-Dipolar cycloaddition of the nitron functionality of **13** and the alkene functionality of **8** yields the backbone-to-backbone cyclised peptides **14-16**. The conformation of these structures is such that they are β -turn mimics. They differ in their C3/C5 stereochemistry with discrete conformational differences.

An increasing interest in peptide secondary structure mimics (*i.e.* α -helix, γ - and β -turns) built-up from scaffold-like molecules is notable.¹ Among these secondary structure mimics, considerable attention has been paid to those mimicking β -turns. Thirteen types of β -turns can be distinguished that vary in their dihedral angles ϕ_2 , ψ_2 , ϕ_3 , and ψ_3 (Figure 1).



A mimic of a β -turn should fulfil the following criteria.¹ It should: (i) reproduce the spacial area of a β -turn (ii) contain the side chains of the amino acid residues *i*+1 and *i*+2 in the correct stereochemistry (iii) minimize steric interactions beyond the peptide backbone^{1f} (iv) still contain the N- and C-terminal ends. A backbone-to-backbone cyclisation in a peptide sequence would fulfil these criteria.² This cyclisation might be achieved by replacement of the N-H...C=O H-bridge in **1** by a covalent bond.³ For the formation of this covalent bond we selected the 1,3-dipolar nitron-alkene cycloaddition reaction. The reaction partners **3** and **4** are accessible starting from amino acids and the adduct is the bicyclic, rigid scaffold **2**. Part of this structure is a isoxazolidine moiety having two new chiral centres (C(3) and C(5)). Thus the cycloaddition reaction will give a mixture of stereoisomers representing several types of the β -turn.⁴

As a touchstone of this approach, we set out to prepare the RGDF β -turn mimics⁵ **14-16**, all sharing the scaffold **2**. Alkylation of **5** with the allylbromides **6** was performed under *phase transfer conditions* (PTC) resulting in the desired mono-alkylated products **7** as main product (Scheme 1). As side-product, some bis-alkylated amine was isolated (5-10%). Several of the established methods for peptide coupling failed to give **8** on reaction of **7** with Z-Asp(OtBu)-OH.⁶ The method of choice was found to be the addition of two

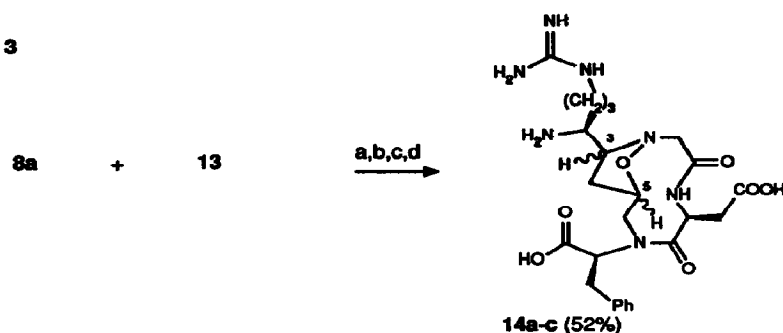


equivalents of DCC as well as Z-Asp(OtBu)-OH to **7a** (**7b**).

$\text{Boc-Arg(Z)}_2\text{-OH}$ (**9a**)⁷ was transformed into the corresponding argininal derivative **10** via the methyl ester **9b** (Scheme 2). The aldehyde was not isolated as such,⁸⁻¹⁰ but immediately treated with N-hydroxyglycine (**12**), obtained from compound **11**.¹¹ We assume that the nitron moiety of the dipeptide **13** has a Z -configuration.¹² 1,3-Dipolar cycloaddition of the nitron functionality of **13** and the alkene moiety of **8a** under high pressure conditions¹³ gave the backbone-to-backbone linkage (Scheme 3). Subsequently, compound **14** was prepared by the following three steps: reductive removal of the protective groups Z and Bzl , amide bond formation and finally removal of the protective groups Boc and tBu . Preparative HPLC separation gave three compounds **14** in a ratio of $\text{14a/14b/14c} = 1/1/3$. These compounds differ in the stereochemistry at the atom $\text{C}(3)$ and $\text{C}(5)$; The NMR spectra were too complex to allow an assignment of the relative stereochemistry.¹⁴

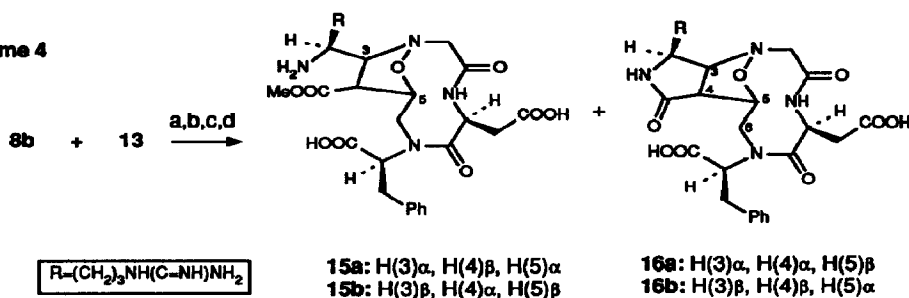
In an analogous fashion, the alkene **8b** was subjected to the nitron cycloaddition reaction. Surprisingly, a mixture of the two diastereomers **15a** and **15b** (ratio 1:1) and the two tricyclic diastereomers **16a** and **16b** (ratio 2:1) was isolated in an overall yield of 49% (Scheme 4).¹⁵ Apparently, an additional cyclisation had occurred in those diastereomers of **15** of which the protons of $\text{C}(3)$ and $\text{C}(4)$ of the isoxazolidine moiety have a *cis*-relationship, resulting in the tricyclic compounds **16a,b**. NOE measurements on the protons

Scheme 3



a) toluene, 15 kbar, 50°C, 2 days b) MeOH, Pd/C, H₂ c) DMF (0.0001M), TBTU (1 equiv.)
pH 8.0, RT d) TFA/phenol/H₂O/HSi(i-C₃H₇)₃ (88/5/5/2)

Scheme 4



a) toluene, 15 kbar, 50°C, 2 days b) MeOH, Pd/C, H₂ c) DMF (0.0001M), TBTU
(1 equiv.), pH 8.0, RT d) TFA/phenol/H₂O/HSi(i-C₃H₇)₃ (88/5/5/2)

H α (Arg), H3, H4 and H5 could discriminate between compounds 16a and 16b. For both compounds, cross-peaks (NOE) were observed between the protons H3/H4; in the NOESY spectra no cross-peak was observed between H3/H5. For 16a, an additional cross-peak between the protons H3/H α (Arg) was present. For the bicyclic compounds 15a,b, NOE experiments failed to give conclusive evidence regarding the configuration of the carbon atoms C(3) and C(5).

None of the compounds 14-16 showed inhibitory activity in the ADP-induced human platelet aggregation assay (GP IIb/IIIa receptor).¹⁶ This finding suggests that the bioactive conformation of RGD required for the GP IIb/IIIa receptor is not mimicked by the β -turns 14-16.¹⁷

However, we demonstrated that the 1,3-dipolar cycloaddition as method for the backbone-to-backbone cyclisation provides several stereoisomers of a relatively rigid, scaffold-based molecule, allowing the study of the bioactive conformation of a given peptide. In the present study, we focussed on the β -turn motif; however, we expect that the 1,3-dipolar cycloaddition as a method for backbone-to-backbone cyclisation might also be applicable to generate mimics featuring a γ -turn or an α -helical scaffold.

References and Notes

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14. The purity of 14a-c (HPLC) was between 90 and 95%; FABMS 534 (M+H), 532 (M-H)
15. 15a-b: HPLC purities were between 80 and 90%; FABMS 591 (M+H), 589 (M-H); The NMR spectra were to complex to allow an assignment of the relative stereochemistry.
 16a: HPLC 96%; FABMS (%) 560 ([M+H]⁺, 22), 558 ([M-H]⁺, 11); δ (D₂O) 7.42-7.27 (m, 5H, C₆H₅), 4.90 (dd, 1H, J=5.6 Hz and J=9.0 Hz, C₆H(Phe)), 4.65 (d, 1H, C₆H(Asp), J=11.0 Hz), 4.24 (dd, 1H, H(3), J_{3,4}=6.2 Hz, J_{3,OH(Arg)}=4.0 Hz), 3.99 (dd, 1H, H(5), J_{4,5}<0.3 Hz, J_{5,6}=4.4 Hz, J_{5,6'}=11.0 Hz), 3.81 (m, 1H, C₆H(Arg)), 3.82 (d, 1H, CH_A(Gly), ²J=14.0 Hz), 3.36 (m, 1H, H(6')), 3.26 (m, 1H, H(6)), 3.25 (d, 1H, CH_B(Gly), ²J=14.0 Hz), 3.25-3.19 (m, 2H, C₆H₂(Arg)), 3.16 (d, 1H, H(4), J_{4,3}=6.2 Hz), 2.82 (dd, 1H, C₆H_A(Asp), J=11.0 Hz, ²J=16.5 Hz), 2.78 (dd, 1H, C₆H_A(Phe), J=9.0 Hz, ²J=15.0 Hz), 2.41 (dd, 1H, C₆H_B(Phe), J=5.6 Hz, ²J=15.0 Hz), 2.01 (br d, 1H, C₆H_B(Asp), ²J=16.5 Hz), 1.74-1.59 (m, 4H, C₆H₂C₆H₂(Arg))
 16b: HPLC 97.3%; FABMS 560 (M+H), 558 (M-H); δ (D₂O) 7.45-7.32 (m, 5H, C₆H₅), 5.47 (d, 1H, J=1.0 Hz), 5.13 (t, 1H, H(3), ³J_{3,4}=8.0 Hz), 4.32 (dt, 1H, C₆H(Asp), J=1.0 Hz, J=4.0 Hz), 4.27 (dt, 1H, H(5), J_{5,4}=11.5 Hz, J_{5,6}=2.0 Hz, J_{5,6'}=9.5 Hz), 4.05 (m, 1H, C₆H(Arg)), 3.99 (t, 1H, C₆H(Phe), J=7.0 Hz), 3.49 (dd, 1H, H(6), J_{6,5}=2.0 Hz, ²J_{6,6'}=13.0 Hz), 3.34 (dd, 1H, H(6'), J_{6,5}=9.5 Hz, ²J_{6,6'}=13.0 Hz), 3.26 (d, 2H, C₆H₂(Phe)), J=7.0 Hz), 3.27-3.15 (m, 2H, C₆H₂(Arg)), 2.91 (dd, 1H, C₆H_A(Asp), J=4.0 Hz, ²J=16.5 Hz), 2.65 (dd, 1H, H(4), J_{4,3}=8.0 Hz, J_{4,5}=11.5 Hz), 2.62 (dd, 1H, C₆H_B(Asp), J=4.0 Hz, ²J=16.5 Hz), 1.85-1.50 (m, 4H, C₆H₂C₆H₂(Arg)).
16. Activation of platelets by ADP is mediated by the GP IIb/IIIa receptor.
17. Upon completion of these studies, this suggestion was corroborated by the finding that the bioactive conformation for this receptor has a turn at Arg, extended ϕ , ψ angles at Gly and a γ -turn at Asp, see; Ku, T.W.; Ali, F.E.; Barton, L.S.; Bean, J.W. *et.al. J.Am.Chem.Soc.* **1993**, *115*, 8861 and references 13-15 therein. In retrospect, our choice to test the RGD-turn mimics reported here on the GP IIb/IIIa receptor was unfortunate.

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